

Further Histological and Ultrastructural Observations on the Cestode, *Senga Mastacembeli* Sp. N. with the Revision of Genus Specieses

Samir J. Bilal¹ & Shamall M. A. Abdullah² & Zohair I. F. Rahemo³

¹ Department of Fish Res. Aqu. Animals- College of Agriculture/ Salahaddin University

² Department of Biology- College of Education/ Ishik University

³ Department of Biology- College of Education, Hamdania University, Enkawa, Erbil.

Correspondence: Samir J. Bilal, Animals- College of Agriculture/ Salahaddin University. Iraq.

E-mail: samir.bilal@su.edu.krd

Received: June 5 , 2017

Accepted: August 24, 2017

Online Published: September 1, 2017

doi: 10.23918/eajse.v3i1sip27

Abstract: A total of 122 *Mastacembelus mastacembelus* were collected from Greater Zab river near Erbil city, Kurdistan region, north of Iraq, during the period from January to the end of December 2011. In the laboratory, the fish was identified and then the fishes were opened from the abdominal site and the gastrointestinal tract was removed out and examined carefully for cestodes. Live cestodes were recovered from the intestine of the infected fish and examined by compound light microscope and scanning microscope, also histological sections were prepared for some samples. The study showed a number of differential characters of *Senga mastacembli* sp. n. which not revealed during the first description by Rahemo (1996) also during the ultra-structural study of it by Rahemo and Mohammad. (1998). The study improved that the species *S. Mastacembli* is a valid species name, also the type host of it *M. Simach* is regarded as a synonyme to *M. mastacembli* and the last is regarded as a type host for this cestode.

1. Introduction

The genus *Senga* was established with the type species *S. besnardi* from Siamese fighting fish, *Betta splendens* in an aquarium at Vinecunes, France (Dolfus, 1934). Cestodes of the genus *Senga* (Order: Pseudophyllidea; Family: Bothriocephalidae) are parasitizing the intestine of freshwater teleostes (Schmidt, 1986). In members of the genus *Senga* hooks on scolex arranged in two semicircles, bothria which are shallow and well developed, apical disc intended dorsally and ventrally. Neck absent. External metamerism present but incomplete. Proglotids acraspedote, wider than long except when gravid. Genital pores dorsal, virtually median. Testes medullary, in two lateral fields. Ovary compact, median, posterior. Vitelline follicles cortical, continuous around margins of proglottid. Uterus loops forward, uterine sac opens ventrally by median pore near anterior margin of proglottid. Eggs operculate (Khalil et al., 1994).

According to Yamaguti (1959), four species of *Senga* reported from different species of fishes in the world. While, eight species have been recorded from Singapore (Polyakova and Kirin, 2005),

recently 27 species have been recorded from India (Bhure and Nanware, 2011). In Iraq, only one species recorded namely *S. mastacembeli* Rahemo, 1996 which described as a new species by from *Mastacembelus simach* collected from Tigris river near Musol city (Mhaisen, 2012).

The present study was planned for redescription of *S. mastacembeli*, including its morphology, surface ultra structure and histologic structure which parasitizes *M. mastacembelus* in the Greater Zab River and brief comparison of the genus *Senga* with its nearest genus, *Polygonchobothrium*.

2. Materials and Methods

A total of 122 *Mastacembelus mastacembelus* were collected from Greater Zab river (tributaries of the Tigris river) at Gwer district near Erbil city, Kurdistan region, north of Iraq, during the period from January to the end of December 2011. The fish specimens were collected by gill netting and cast netting by local commercial fishermen.

In the laboratory, the fish was identified according to Coad (2010); Froese and Pauly (2016), and then the fishes were opened from the abdominal site and the gastrointestinal tract was removed out from the rectum to the esophagus and opened longitudinally and examined carefully for cestodes (Amlacher, 1970; Barson, 2004).

For light microscopy (LM), a live cestodes were recovered from the intestine of the infected fish. The parasites were then washed in 0.6% saline solution, fixed in hot 4% formalin and preserved in 70% ethanol. The staining is accomplished by using Schuberg's iron hydrochloric carmine by Regressive staining method, destaining was performed with an acid ethanol (HCl/ETOH) solution and stopped by transferring the samples into 80% ethanol. Dehydration by using of graduated concentrations of ethanol. The specimens put between a coversliple and a piece of paper to avoid coiling during dehydration, clove oil used for clearing of the specimens after dilution by graduated concentrations of ethanol. Drawings were made with the aid of a Zeiss microscope drawing attachment (Scholz & Hanzelová, 1998). Photos were taken with Olympus camera (Japanese origin), and the figures were drawn by using a camera lucida (drawing tube).

For histological study, the samples of cestodes preserved in 70% ethanol then dehydrated by graduated concentration of ethanol (80%, 90% and 96% ((twice)), then ethonol changed with acetone for 35-40 min., then transfered to toluen for one hour then to a new toluen for 1.30 hour. The specimens embedded in Paraplast media (Paraplast Regular) manufactured by Sigma-ALDRICH co. for 24 hours then transfered to a new Paraplast for 48 hours and third paraplast for 72 hours in oven on 56C°. Then the samples were placed in a perpendicular position under a dissecting microscope, which has to be done quickly before the paraffin hardens. After 30 minutes the vial was placed in the refrigerator, to be become hard in order to remove the paraffin-enclosed sample from the vial.

After embedding, sections where obtained by using microtome at 8-12 µm thick, stretched on 40 C° water bath then fixed on slides with help of 80 C° hotplate. Wax were removed from sections by using xylene then descending sereis of alcohols were used for dehydration. Sections were stained using hematoxyline-eiosin, finally mounted in canada balasm (Scholz & Hanzelová, 1998).

Specimens for the Scanning Electrone Microscopy (SEM) were fixed and preserved like that used for (LM), then dehydrated completely. Chemical method, Hexamethyldisilazan was used for drying of tape worms by covering the specimens with this material for 5-10 min., then the chemical taked out totally with a droper, leave the worms to dry at room temp. Finally, when the specimens totally dried

embedded on the target and sputter-coated with gold; they were examined using a JEOL JSM-7401F scanning electron microscope (Academy of Czech Republic - parasitology institute) at an accelerating voltage of 4 kV GB low linked to an external computer system. Each specimen was observed specially on the morphology of the scolex (Scholz & Hanzelová, 1998).

3. Results and Discussion

Fishes were surveyed for parasitic cestodes during the period of the present study. The survey showed the occurrence of one cestode belonging to the genus *Senga*.

Senga mastacembeli Rahemo, 1996

Host: *Mastacembelus mastacembelus* (Marmarej).

Prevalence of infection: 73.77%.

Mean intensity: 2.3.

Site infection: Intestine.

Locality: Greater Zab river at Gwer District.

Description: Medium sized, white yellowish worms. Body length 90-123 mm, width 2.12-2.89 mm. Scolex lancet shaped, 0.561- 0.752 mm length, 0.865-0.917 mm width (Fig. 1A; 2A; 4A). Bothria 0.653-0.739 mm length, 0.174-0.189 mm width (Fig. 1B; 1C; 2B; 4B). Rostellum 0.083-0.91 mm length, 0.168-0.183 mm width (Fig. 1B; 2B; 4B). Hooks are in one row and two semi-circles around the anterior end of rostellum, 48-50 in number, 0.023-0.031 mm length, 0.0052-0.0078 mm width (Fig. 1C; 2B; 4C; 4D; 4E). Immature proglotid, 0.351-0.405 mm length, 0.462-0.492 mm width (Fig. 1D). Mature proglotid wider than long, 0.357-0.374 mm length, 1.298-1.342 mm width (Fig. 1E; 1F; 2C). The ovary is bilobed and compact in texture, 0.299-0.345 mm in length and 0.065-0.079 mm in width (Fig. 1E; 2C), similar observations were recorded by Rahemo (1996). Oviduct, 0.059-0.065 mm length (Fig. 1E; 2C). Uterus highly curved, 0.389-0.412 mm length, 0.023-0.034 mm width (Fig. 1E; 2C). Vagina median in ventral surface of the proglotid, 0.034-0.041 mm length, 0.132-0.139 mm width (Fig. 1F; 2C). Cirrus sac semi spherical to sub spherical, alternated in proglotids measured, 0.174-0.183 mm length, 0.213-0.233 mm width and with a thick muscular wall (Fig. 1E; 2C). The testes in the present specimens varies between 65 -72 in mature segments, situated at the lateral fields, each 0.0132 -0.167 mm length, 0.0226-0.0261 mm width not reaching the ovary level, and are medullary in position (Fig 1E; 2C; 3C; 3D). Anyhow, the number of testes reported in the present specimens is less than that estimated by Rahemo (1996) it may be due to maturation status, but in both studies testes are found in medulla. As seen in (Fig 1E; 1F; 3A; 3B; 3C; 3D), most of the reproductive organs are demonstrated either grossly or in both longitudinal and cross sections. Vitelline follicles 0.0089-0.0097 mm length, 0.0078-0.0091 mm width (Fig 1F; 2C). The vitelline follicles in the examined specimens lie on both sides of the mature segments occupying extensive area (Fig 1E; 1F; 1G). Furthermore, these vitelline follicles are cortico-medullary in position but more abundant at the medulla. Such results were not reported by Rahemo (1996) and Rahemo and Mohammad (1998).

Gravid segment, 0.448-0.477 mm length, 1.821-2.45 mm width, containing an enlarged sac like uterus, 0.356-0.371 mm length, 0.734-0.762 mm width, filled with eggs (Fig. 1G; 2D; 3C). Uterus is

filled with eggs which are operculated, oval in shape 0.0023-0.0031 mm length, 0.0012-0.0013 mm width, the operculum was not seen by Rahemo (1996) and Rahemo and Mohammad (1998). Further studies are suggested to explore the factors influencing the hatching of these eggs i.e. the removal of this operculum and the emerging of the oncomiracidium of the present species which was not observed neither in this study nor in the previous studies (Rahemo, 1996; Rahemo and Mohammad, 1998). However, experimental studies may reveal the stages of the life cycle of the present worm both in Greater Zab river from which the present fishes were collected or from Tigris river from which Rahemo's specimens were collected.

Scanning electron microscopy of the scolex showed the presence of fine, hairy microtrichia, medium in length about 10 μm (Fig. 4F), these microtrichia usually present in cestodes (Roberts and Janovy, 2005) their function is not known but it may help in the attachment of the worms to intestinal lining. These microtrichia were not observed by Rahemo (1996) as he did not use SEM. The microtrichia of the scolex (Fig. 4E) seem to be irregular, some are short others are long, some posteriorly directed while others are transversely directed. Comparing the microtrichia of this worm with *Hymenolepis nana*, in the latter they are all posteriorly directed while in other cestodes, *Trypanorhynch* they are five to six-fingered palmate microtrichia (Roberts and Janovy, 2005). Also (Fig. 4F) show hard and well formed hooks which are posteriorly curved and pointed distally while their proximal part are inserted in the rostellum. Furthermore, these hooks are arranged in two semicircles (Fig. 4B; 4C; 4E), these observations were not noticed by Rahemo (1996). In addition, scanning electron micrographs show that the bothria are deep, highly muscled and with glandular tips (Fig. 4C; 4D). Anyhow, the glandular nature of these glandular tips are to be confirmed by using transmission electron microscopy in order to have prediction about their function, which is likely a secretion used for attachment. As in (Fig. 4F) these microtrichia are of filamentous and canoid type which directed posteriorly similar to the observations of transmission electron microscope observation carried out by Rahemo (1998). Generally these microtrichia are similar to microvilli present in the gut mucosa of vertebrates and invertebrates. Electron microscopic study show that each microtrichia have a dense distal portion set off from the base by a multilaminar plate (Roberts and Janovy, 2005). Anyhow, Rahemo and Mohammad (2002) made a scanning electron microscopy for the scolex of this cestode but their observation were oblique and incomplete as the microscope at that time was not working properly. Neck absent (Fig. 4B).

As revealed from both longitudinal and cross sections (Fig. 3A; 3B; 3C; 3D) there are no septa between proglottids which is a general style of cestode structure, such phenomenon was not recorded in the previous studies (Rahemo, 1996; Rahemo and Mohammad, 1998).

From the paraffin sections musculature not developed highly, only some longitudinal and transverse bundles were located underneath the tegument (Fig. 3C; 3D). The vitelline follicles are cortical, compact and spherical in transverse sections (Fig. 3C; 3D). Testes are medullary, arranged in two incomplete rows, starting from the uterus anterior end level and extended to the end of posterior end level of ovaries (Fig. 3A; 3C; 3D). Ovary is bilobed, compact, connected by narrow connections with short oviducts (Fig. 3C; 3D). The most important observation in the present study is the Mehlis' gland which is found to be large, spherical or subovoid in cross section lie just directly beneath the ovary loops, sometimes may pass to other adjacent proglottids since there are no intersegmental septa in the present worm (Fig. 3A; 3C).

It is noteworthy that most of the above observations were lacking in examination of the Senga

specimens carried out more than three decade ago.

The present study regarded as a first redescription of this species in the world including a lot of important characters like shape and position of vettilaria and testes, shape and long of oviduct, Mehlis' gland, vagina and eggs, also it is histology and surface ultra structure (like michrotriches) characters by using scanning electron microscope. The fish host of the present cestode is *M. mastacemulus*, while in the previous studies, *Mastacembelus simach*, more likely these two names are synonemous according to fishbase (2011) and Coad (2010), in his valid book gave no report of *S. simach*.

Bashě and Abdullah (2010) described *Polyonchobothrium magnum* from the intestine of the same host (*M. mastacemulus*) and locality (Greater Zab river) with very similar characters and hooks arranged in two semicircles and since the hooks of *Polyonchobothrium* spp. are arranged in four quadrants, for this, this name regarded as a synonyme for *S. mastacembeli* (Khalil et al., 1994).

Species revesion of genus *Senga*:

According to Bhure and Nanware (2011) there are 26 other species under this genus were recorded from freshwater fishes namely:

Senga ophiocephalina from *Ophiocephalus argus* at Taimen, China.

S. lucknowensis from *Mastacembelus armatus* in India.

S. malayana from *Channa striata*.

S. parva and *S. filiformis* from *Channa micropeltes* at Malacca.

S. pahangensis from *Channa micropeltes* at Tesak Bera.

S. besnardi from *Ophiocephalus gachua* in India.

S. visakhapatanamensis from *Ophiocephalus punctatus* in India.

S. khami from *Ophicephalus marulius* from Kham river at Aurangabad.

S. godavari from *M. armatus* at Nanded, M.S. India.

S. aurangabadensis was added from *M. armatus* at Aurangabad M.S. India.

S. paithaniensis from *M. armatus*.

S. raoi and *S. jagannathae* from *Channa punctatus*.

S. maharashtrii and *S. gachuae* from the intestine of *M. armatus*.

S. chauhani from *Channa punctatus*.

S. mohekarae from the intestine of the *M. armatus*, at Parli, Dist. Beed, M.S. India, *S. armatusae* from *Mastacembalus armatus*.

S. tappi from *M. armatus*.

S. ayodhensis from *Amphinuous cuchia*.

S. baghui from *Rita rita*.

S. jadhavae from *Mastacembelus armatus*.

S. chandapurensis from *Mastacembelus armatus*.

S. ticto from *Punctatus ticto*.

S. madhavae from *M. armatus*.

Rego (1997) described a new species of cestodes belonged to the genus *Senga* from the intestine of *Astyanax scabripinnis* from Brazil namely: *S. besnardi*. The distinguishing features of this species are scolex rectangular, with an apical bilobed disc whose margins carry a row of hooks, divided in two half circles of hooks, each half has 14-15 hooks of different size and form, some vestigial and rudimentary hooks intercalated between the others, as described to *S. besnardi*

Polyakova and Kirin (2005) described *Senga sharpiloi* from *Channa micropeltes* from Singapore. *S. sharpiloi* differs from others species of genus *Senga* by the size strobili length, scolex is pear shaped, apical disc elongated slightly, hooks of the ends of both semicircles are smaller than the others and number of hooks (44-50 + 4), number of testes which is 22-24 in each proglottid and arrangement of them.

Bhure and Nanware (2011) described *S. satarensis* from *M. armatus* at various places of India. The worm comes closer to all the known species of this genus in general topography of organ but differs due to scolex pear shaped, tapering anteriorly and broad posteriorly, rostellum medium, rounded, bearing 28-30 rostellar hooks, mature proglottids six to seven times broader than long, testes 175-200 in numbers, scattered throughout the segment, cirrus pouch oval, ovary distinctly bilobed, vagina thin, runs posteriorly, genital pores oval in shape, vitellaria granular, uterus saccular and egg elongated.

Wankhede et al. (2012) studied the histopathological changes of the intestine of freshwater fish *M. armatus* infected with *Senga* sp. and noticed destruction and extrusion of the intestinal villi, fibroblast cells and plasma cells.

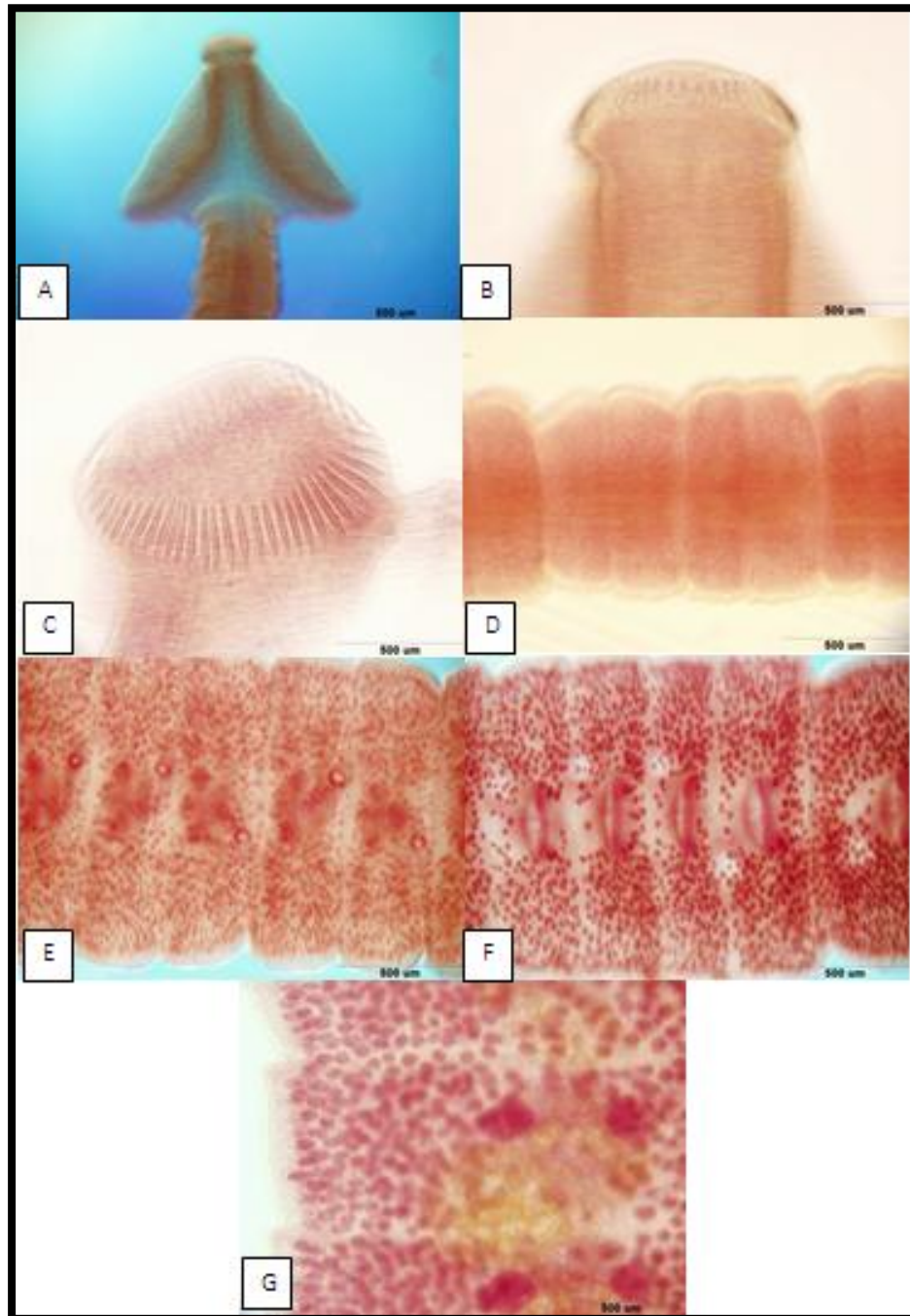


Figure (1): Photomicrograph of *Senga mastacembeli*.

A- Scolex, B- Rostellum, C-Pressed proscelis show hooks, D- Immature proglotid, E- Mature proglotid (Dorsal view), F- Mature proglotid (Ventral view), G- Gravid proglotid.

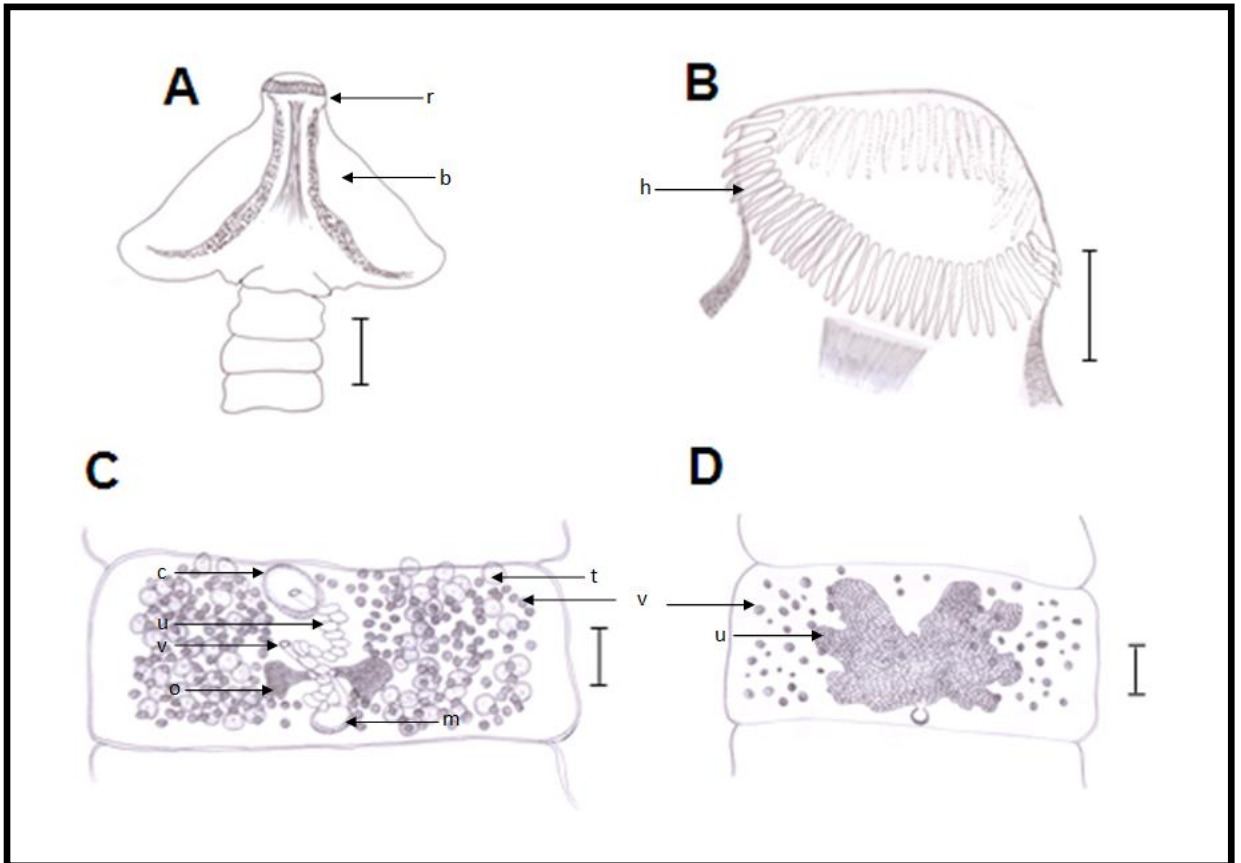


Figure (2): Lucida drawings of *S. mastacembeli*.

A- Scolex, B- Rostellum, C- Mature proglotid, D- Gravid proglotid.

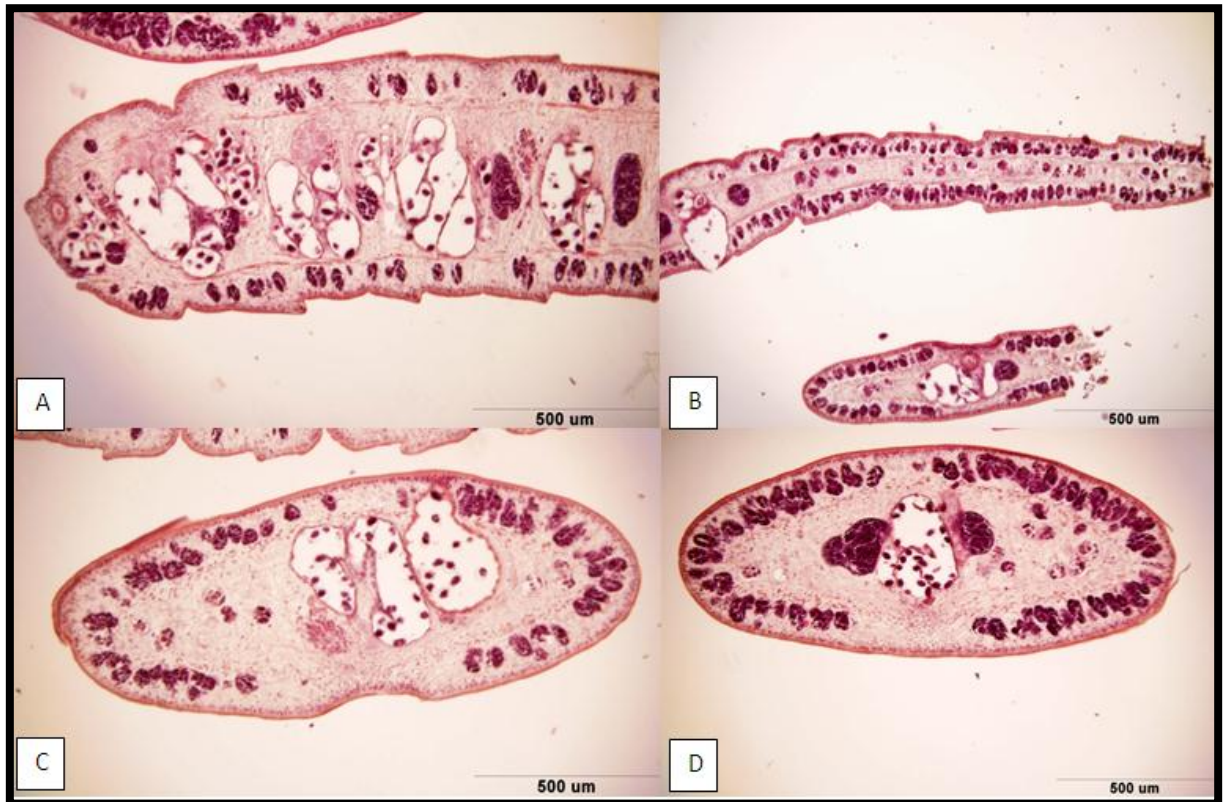


Figure (3): Photomicrograph of *S. mastacembeli* sections.

Longitudinal section of gravid proglotids, B- Longitudinal section of mature proglotids and cross section of mature proglotid through mehl's gland, C- Cross section of mature proglotid through pre-ovarian region, D- Cross section of mature proglotid through ovarian region.

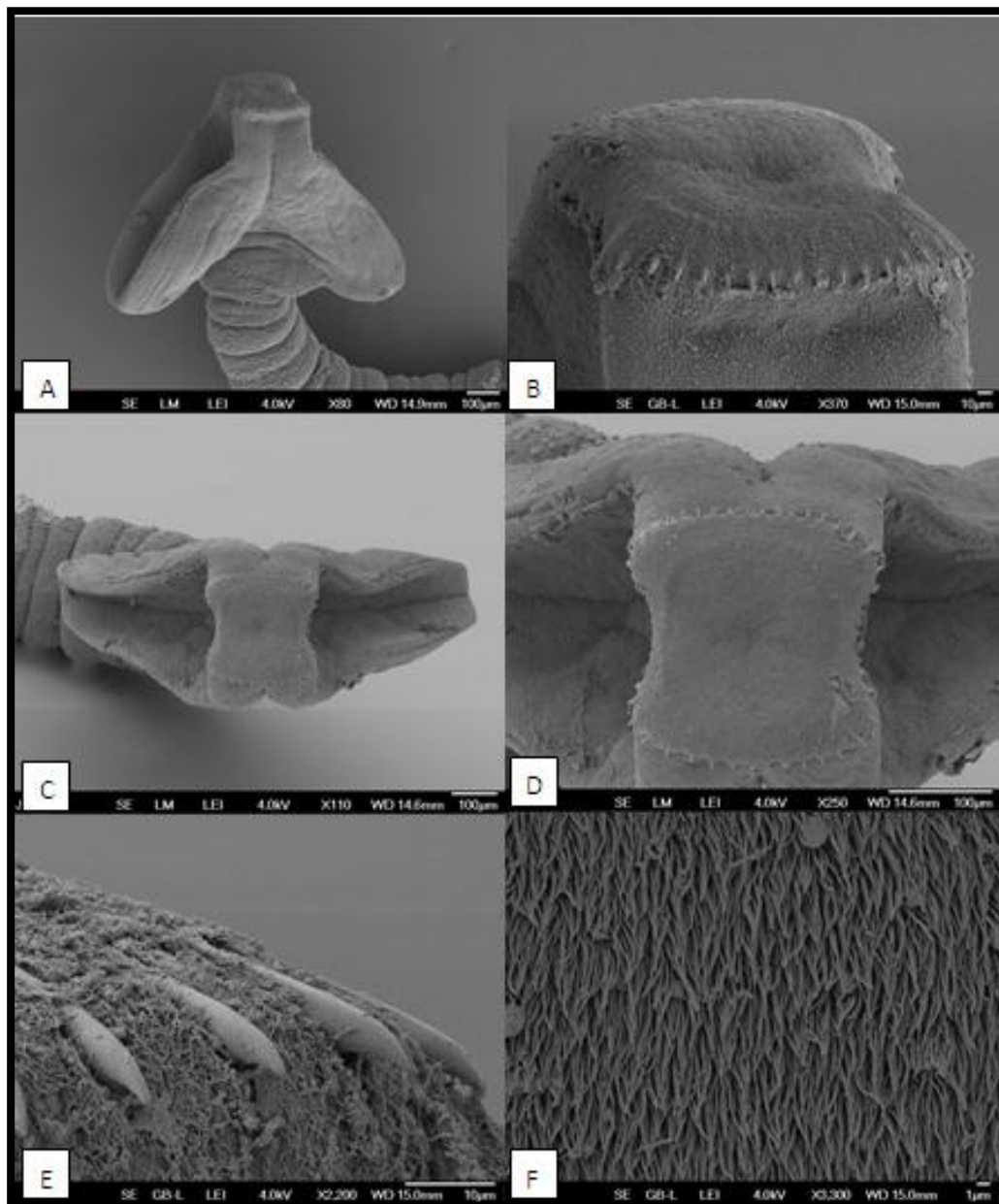


Figure (4): Scanning electron micrograph of *S. mastacembei*.

Scolex (Dorsal view), B- Rostellum (Dorsal view), C- Scolex (En face view), D- Scolex (En face view) showing the hooks hemispheres, E- Hooks, F- Microtriches from proglotid surface.

References

- Amlacher, E. (1970). Textbook of fish diseases (Engl. Transl.). T.F.H. Publ., Jersey City: 302pp.
- Barson, M. (2004). Endoparasites of the sharptooth catfish, *Clarias garirpinus* (Burchell), from the Rietvlei dam, Sesmyl Spirit system, South Africa. M. Sc. Thesis, Coll. Sci., Univ. Rand Afrikaans: 53pp.
- Bashē, S. K. R. & Abdullah, S. M. A. (2010). Parasitic fauna of spiny eel *Mastacemelus mastacemelus* from Greater Zab river in Iraq. *Iranian J. Veter. Res., Shiraz Univ.*, 11, (1), 30, 18-27.

- Beckman, W. C. (1962). The freshwater fishes of Syria and their general biology and management. FAO Fish. Biol. Branch Tech. Pap. No.8: 179pp.
- Bhure, D. B. & Nanware, S. S. (2011). Systematic observation of new pseudophyllidean tapeworm Senga [1] from *Mastacembelus armatus*. *Inter. Multidisciplinary Res. J.*, 1(10), 25-28.
- Coad, B. W. (2010). Freshwater fishes of Iraq. Pensoft. P., 258-259.
- Dollfus, R. Ph.(1934). Sur uncestode pseudophyllidae parasite de poisson ornement. *Bull. Sac. Zool.*, 69, 476-490.
- Froese, R. & Pauly, D. (2016). FishBase. World web electronic publication. www.fishbase.org, version (11/2016).
- Khalil, L. F.; Jones, A. & Bray, R. A. (1994). Keys to the cestode parasites of vertebrates. CAB Intern., Wallingford, UK. 751 pp.
- Nepszy, S. J. (1988). Parasites of fishes in the Canadian waters of the Great Lakes. Great Lakes Fish. Comm. Tech. Rep. No. 51: 106pp.
- Polyakova, T. A. & Kirin, M. P. (2005). *Senga sharpiloi* sp. nov. (Cestoda: Ptychobothriidae) a new cestode species from *Channa micropeltes* (Pisces: Channidae) from Singapore. *Ecol. Sea*, 68, 68-75.
- Rahemo, Z. I. F. (1996). Parasites of the Iraqi spinyback *Mastacembelus simach* (Walbaum, 1792). *Iraqi J. Biol. Sci.*, 15, 42-52.
- Rahemo, Z. I. F. & Mohammad. (1998). Ultrastructural studies of the piscin cestod, *Senga mastacembeli* Rahemo, 1992. *Riv. Parasitol.*, XV (LIX) 1: 59-66.
- Rahemo, Z. I. F. & Mohammad. (2002).
- Rego, A. A.(1997). *Senga* sp., Occurrence of a Pseudophyllid Cestode in a Brazilian Freshwater Fish. *Mem. Inst. Oswaldo Cruz*, 92(5): 607.
- Roberts, L. S.; & Janovy, J. (2005). Genrald, D. Schmidt and Larry S. Robert's Foundations of parasitology, 7th edn. McGraw- Hill Co., Inc., New York: 702pp.
- Scholz, T. & Hanzelová, V. (1998). Tapeworms of the Genus *Proteocephalus* Weinland, 1858 (Cestoda: Proteocephalidae), Parasites of Fishes in Europe. *Studie AV ČR, Academia Praha 2* (98): 119 pp.
- Schmidt, G. D. (1970). How to know the tapeworms. Wm. C. Brown Co. Publ., Dubuque: 266pp.
- Wankhede, H. J.; Aade, U. P. & Kaldate, K. D. (2012). Histopathological study of Ptychobothriidan tapeworm in freshwater fish *Mastacembelus armatus* (Lecepede, 1800) from Aurangabad district (M. S.). *Indian Streams Res. J.*, 2(2), 1-4.